

S.M. Barmak^{1,2}, Yu.A. Sinyavskiy², A.B. Berdygaliev², I.S. Savitskaya¹,
T.Sh. Sharmanov¹, I.H. Mendenhall³, E.V. Zholdybaeva⁴

¹Al-Farabi Kazakh National University, Almaty, Kazakhstan;

²Kazakh Academy of Nutrition, Almaty, Kazakhstan;

³Duke-NUS Medical School, Singapore;

⁴National Center for Biotechnology, Nur-Sultan, Kazakhstan

(E-mail: sabyr95@mail.ru)

Selection of an effective DNA extraction method for the detection of *Salmonella* by the polymerase chain reaction

DNA extraction is a crucial step at using molecular and biological methods, since the result of PCR analysis largely depends on the quality and quantity of the obtained DNA preparations. In this work, we compared the efficiency of DNA extraction from bacteria in various ways: the trizol method using TRizol commercial reagent, the CTAB method and the sorbent method using silica particles. It has been shown that the extraction of DNA from *Salmonella* bacteria on silica particles ensures higher sensitivity than the other methods. The amplification of the extracted DNA on silica particles is more efficient and allows obtaining reproducible results with the accuracy of a high degree. In the future, this method will be used to complete the test system for diagnosing salmonellosis. Introduction of the methods of express diagnosis into practice is extremely important, as quick and early diagnosis will allow counteracting the spread of infection in a timely manner, which in turn will improve the epidemiological and epizootiological situation in the country.

Keywords: *Salmonella* bacteria, method, identification, DNA extraction, polymerase chain reaction, amplification, silica particles.

Introduction

Salmonellosis is a widely spread human and animal infection caused by various members of the *Salmonella* genus. Food raw material and foodstuff are the most dangerous for people of *Salmonella* reservoir. In many European countries, salmonellosis morbidity among humans has significantly increased for two recent decades [1].

Lately, methods of gene diagnosis (nucleic acid hybridization, polymerase chain reaction — PCR, and so on) are successfully used to detect and identify microorganisms in various objects under study. These methods allow indicating and identifying microorganisms at high specificity in the presence of attendant microflora and with sensitivity right up to single cells [2].

The identification of infectious agents by classical bacteriological methods is associated with the usage of numerous selective nutritive media and various substrates. It makes the identification process protracted and laborious. Moreover, growing contaminating microorganisms in various media under competition conditions not always results in adequate identification. Furthermore, *in vivo* identification of transformed bacteria or of artificially genetically changed forms carrying pathogenicity genes not typical for the strain is difficult in the classical bacteriological assay [3].

Currently, there are various methods that allow extracting nucleic acids from a wide range of samples but only a bit of them is suitable for automation and many steps of isolation involve the risk of contamination. The presence of contaminants (proteins, for instance) in the complex mixtures often impedes the implementation of necessary reactions and techniques. Methods of DNA or RNA extraction should meet the following priority requirements: lysis of the biological material, selective extraction (sorption), concentration from large volumes, separation of components that inhibit PCR, separation of DNA and RNA, high yield percentage, opportunity of calibration and of positive control, absence of contamination, low time expenses, automation possibility [4].

Dynamic of development of veterinary and sanitary expert examination and of foodstuff certification is characterized by transfer to the most specific, sensitive and accelerated methods of bacteria identification [5].

Polymerase chain reaction (PCR) has a number of advantages as compared to the traditional bacteriological method of *Salmonella* detection since it combines rapidness and simplicity of implementation as well as potentially high specificity and sensitivity in revealing pathogenic microorganisms [6].

The objective of our studies was to select an effective method of DNA extraction for *Salmonella* detection by the method of polymerase chain reaction.

Materials and Methods

The objects of the study were eight *Salmonella* bacteria isolated from the foodstuff at the Microbiology Chair of Al-Farabi Kazakh National University.

DNA Extraction. DNA was extracted from *Salmonella* bacteria by the following methods:

1. Trizol method. The commercial reagent TRizol («Invitrogen», USA) was used in accordance with the manufacturer's manual.

2. CTAB method. CTAB (cetyl trimethyl ammonium bromide) is a classical cation detergent that is used in DNA extraction. CTAB lyses the cellular membrane, effectively destroys DNA-protein [7].

3. Sorbent method using silica particles. DNA binds to silica particles under the effect of high concentrated salt [8].

The quality and quantity of the extracted DNA were controlled with the use of NanoDrop 2000.

For setting PCR used the same amount of DNA (~ 30 ng/μL) obtained by different methods.

PCR-amplification. PCR-fragments of Inv gene from *Salmonella* bacteria were produced with the use of the following primers: S Inv-1F (direct) and S Inv-1R (inverse). Amplification was carried out in 25 μL of reaction mixture of the following composition: 10 xbuffer of DNA polymerase — 2.5 μL, 10 mM dNTP — 1 μL, MgCl₂ — 1 μL, 20–50 ng of DNA-matrix, 20 pM of direct and 20 pM of inverse primers, and 0.5 U of DNA polymerase (Invitrogen), DNA — 1 μL. PCR products were assayed by electrophoresis in 1.5 % agarose gel containing ethidium bromide (1 μg/μL) at the field strength 6 V/cm². The PCR product sized 500 bp.

Results and Discussion

Comparative study of three different versions of the DNA extraction from *Salmonella* bacteria was carried out: the trizol method using TRizol commercial reagent, the CTAB method and the sorbent method using silica particles.

The parameters such as quantity and quality of the obtained DNA were taken into consideration to select the DNA extraction method. The results of extracting genome DNA by different methods from 8 *Salmonella* isolates are shown in Figure 1.

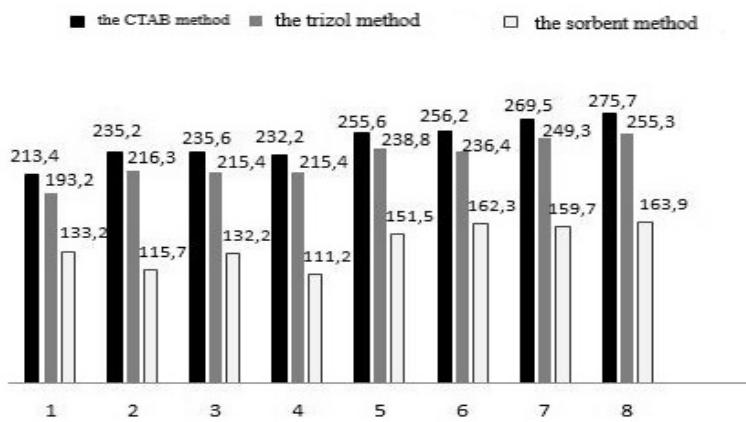
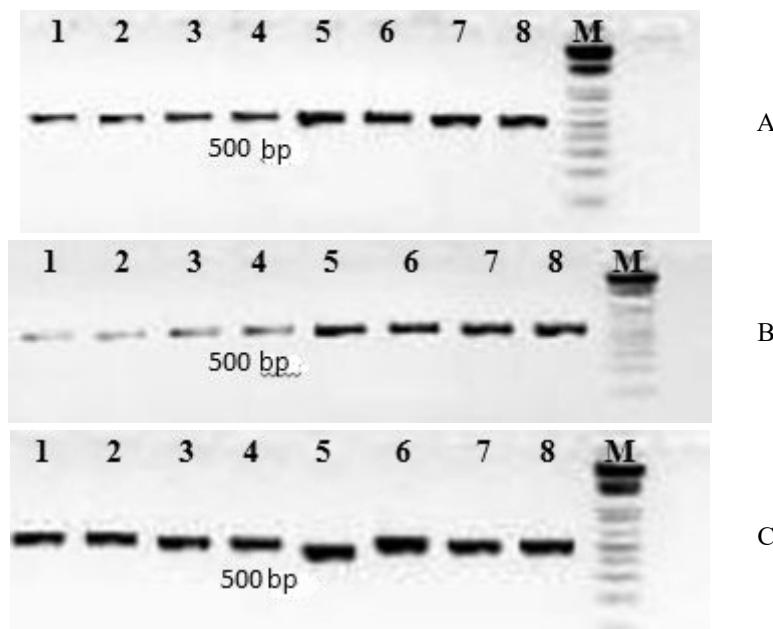


Figure 1. The amount of DNAs extracted from eight *Salmonella* isolates (ng/μL) by different methods

The implemented studies showed the CTAB method to demonstrate the highest DNA yield. The results of the trizol method are slightly lower. And the lowest DNA yield was demonstrated by the sorbent method.

PCR-assay depends significantly on the quality of the extracted DNA. PCR amplification was conducted with the DNA specimens of equal volume (1 μL) obtained by different extraction methods from samples of the materials under study: the trizol method using TRizol commercial reagent, the CTAB method and the sorbent method using silica particles. The results are shown in Figure 2.



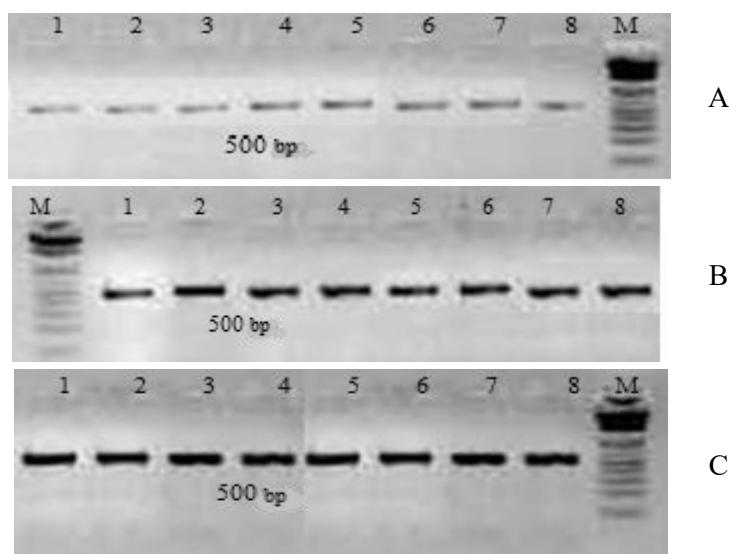
A — TRizol method; B — CTAB method; C — sorbent method;
1–8 — DNAs of *Salmonella* bacteria; M — 1 kb DNA marker (Invitrogen)

Figure 2. Electrophoregram of the DNAs obtained from *Salmonella* bacteria

Figure 2 shows that the genetic material of high quality and of concentration sufficient for PCR-assay has been obtained by the sorbent method with the use of silica particles (C). At the same time, it should be noted that the DNA concentration in these preparations is 1.61 times less than in preparations obtained by using TRizol (A) and 1.75 times lower versus the preparations obtained by the CTAB method.

Insignificant PCR inhibition in the DNA specimens obtained by the trizol method using TRizol commercial reagent and the CTAB method may be caused by the DNA excess, which is a PCR inhibitor.

The same amount of DNA (50 ng/ μ l) was used to select an effective DNA extraction method for detecting *Salmonella* in the PCR reaction. The results are presented in Figure 3.



A — 50 ng/ μ l of DNA isolated with TRizol reagent was used in the PCR reaction; B — 50 ng/ μ l of DNA isolated with CTAB method was used in the PCR reaction; C — 50 ng/ μ l of DNA isolated with sorbent method was used in the PCR reaction; 1–8 — DNAs of *Salmonella* bacteria; M — 1 kb DNA marker (Invitrogen)

Figure 3. PCR products of *Salmonella* bacteria on an agarose gel using 50 ng/ μ l of DNA

As can be seen in Figure 3, optimal results were obtained when PCR was performed using DNA isolated by the sorbent method.

The major criterion in methods of the DNA extraction is that nucleic acid should be the utmost purified from cellular DNA and protein impurities. The extracted genome DNA should be unfragmented since it is a matrix for the synthesis of the specific product [9]. Such a result is ensured by the sorbent method using silica particles. When this method has used the proteins and cellular components are removed and the utmost purified DNA is left. The advantages of the DNA extraction method with the help of silica particles are as follows: loss minimization in the course of the DNA extraction; lower risk of cross-contamination owing to the linkage of all nucleic acid to the sorbent; high purity of the final product [10]. In contrast to other methods, this one does not require the usage of toxic reagents.

Effectiveness of the process of the DNA extraction is a determining factor in the course of work with a small amount of the material or with the specimens containing a considerable quantity of inhibitors that may result in lower diagnostic sensitivity of the test and inhibition of the amplification process [11]. The properly selected method of the DNA extraction allows achieving the most accurate result of the polymerase chain reaction.

Conclusion

On the basis of the implemented study, one can conclude that the sorbent method using silica particles is the most suitable method for DNA extraction from *Salmonella*. In the future, this method may be used to kit up the test system for *Salmonella* identification and genotyping.

The work was carried out within the project of grant financing «Genotyping pathogenic microorganisms in the food raw material and foodstuff realized in the markets and supermarkets of the Republic of Kazakhstan, development of recommendations aimed at reducing the risk of morbidity among the children of preschool and school age», years 2018–2020, No. AP05131147.

References

- 1 Токарев В.А. Варианты мониторинга госпитальных штаммов ВБИ сальмонеллезной этиологии / В.А. Токарев, Е.И. Гудкова, А.А. Адарченко // Внутрибольничные инфекции — проблемы эпидемиологии, клиники, диагностики, лечения и профилактики: тез. докл. Рос. науч.-практ. конф. — М., 1999. — С. 236–237.
- 2 Lim J.K. In Vivo Phase Variation of Escherichia coli Type 1 Fimbrial Genes in Women with Urinary Tract Infection / J.K. Lim, N.W. Gunther, H. Zhao, D.E. Johnson, S.K. Keay, H.L.T. Mobley // Infection and immunity. — 1998. — Vol. 66, No. 7. — P. 3303–3310.
- 3 Абжалиева А.Б. Усовершенствование методов идентификации пищевых патогенов в продуктах животного происхождения: автореф. дис. ... д-ра философии PhD: 6D120200 — «Ветеринарная санитария» / А.Б. Абжалиева. — Алматы, 2017.
- 4 Нургалиева М.Т. Методы выделения нуклеиновых кислот для проведения полимеразной цепной реакции в режиме реального времени / М.Т. Нургалиева, О.О. Жансеркенова, Е.С. Усенбеков, Ш.Н. Касымбекова, А.К. Смагулов // Изденистер, нәтижелер — Исследования, результаты. — 2016. — № 3 (71). — С. 68–72.
- 5 Мамаев М.А. Разработка ускоренных способов ДНК-диагностики для определения бактерий в мясе и мясопродуктах: автореф. дис. ... канд. вет. наук: 16.00.08. — «Гигиена животных, продуктов животноводства и ветеринарно-санитарная экспертиза» / М.А. Мамаев. — М., 2000.
- 6 Herrera-Leo S. Multiplex PCR for Distinguishing the Most Common Phase-1 Flagellar Antigens of *Salmonella* spp. / S. Herrera-Leo, J.R. McQuiston, M.A. Usera // J. Clin. Microbiol. — 2004. — Vol. 42. — P. 2581–2586.
- 7 Scott O. Rogers. Extraction of DNA from milligram amounts of fresh, herbarium and mummified plant tissues / Scott O. Rogers, Arnold J. Bendich // Plant Molecular Biology. — 1985. — Vol. 5. — P. 69–76.
- 8 Капустин Д.В. Высокоэффективный метод одностадийного выделения ДНК для ПЦР-диагностики *Mycobacterium tuberculosis* / Д.В. Капустин, А.И. Простякова, Я.И. Алексеев // Acta naturae. — 2014. — Т. 2, № 21. — С. 52–57.
- 9 Киркимбаева Ж.С. Сравнительное изучение эффективности различных методов выделения ДНК лептоспир / Ж.С. Киркимбаева, С.Е. Ермагамбетова, К.Б. Бияшев // Изденистер, нәтижелер — Исследования, результаты. — 2016. — № 4(72). — С. 50–54.
- 10 Аукенов Н.Е. Выделение и очистка нуклеиновых кислот. Состояние проблемы на современном этапе / Н.Е. Аукенов, М.Р. Масабаева, У.У. Хасanova // Наука и здравоохранение. — 2014. — № 1. — С. 51–53.
- 11 Каталог сайта «ДНК-Технология». — [Электронный ресурс]. — Режим доступа: <https://docplayer.ru/45703435-I-ekspress-metod-vydeleniya-dnk-komplekt-reagentov-dlya-vydeleniya-dnk-proba-rapid-4-komplekt-reagentov-dlya-vydeleniya-dnk-proba-rapid-genetika-5.html>

С.М. Бармак, Ю.А. Синявский, А.Б. Бердыгалиев,
И.С. Савицкая, Т.Ш. Шарманов, И.Х. Менденхалл, Е.В. Жолдыбаева

Полимеразды тізбекті реакция көмегімен сальмонелланы айқындау үшін ДНҚ бөлудің тиімді әдісін таңдау

Молекулярлық-биологиялық әдістерін қолданған кезде ДНҚ бөлу шешуші қадам болып табылады, себебі ПТР-талдауының нәтижесі алынған ДНҚ препараттарының сапасына және санына байланысты болады. Макалада *Salmonella* бактерияларының ДНҚ бөлу тиімділігі түрлі әдістермен салыстырылды: TRizol коммерциялық реагенттің қолданатын тризол әдісі, СТАБ әдісі және кремнезем бөлшектерін қолданатын сорбентті әдіс. ДНҚ кремнезем бөлшектерінде оқшауланған кезде, басқа әдістермен салыстырганда, *Salmonella* бактерияларының ДНҚ-ын анықтау сезімталдығы қамтамасыз етілген. Бөлініп алынған ДНҚ амплификациялануы кремнеземдік бөлшектердің қолданғанда қүштейтілуі тиімдірек және жоғары дәлдіктің қайталанатын нәтижелерін алуға мүмкіндік береді. Болашақта бұл әдіс *Salmonella* диагностикасының сынақ жүйесін жасау үшін қолданылды. Жедел диагностикалау әдістерін енгізу практикада ете маңызды, ейткені инфекцияны тез және ерте диагностика жасау оның таралуына уақытылық қарсы тұруға мүмкіндік береді, бұл, өз кезеңінде, елдегі эпидемиологиялық және эпизоотологиялық жағдайдың деңгейін арттыруды.

Кітт сөздер: *Salmonella* бактериялары, әдіс, айқындау, ДНҚ бөлу, полимеразды тізбекті реакция, амплификация, кремнезем бөлшектері.

С.М. Бармак, Ю.А. Синявский, А.Б. Бердыгалиев,
И.С. Савицкая, Т.Ш. Шарманов, И.Х. Менденхалл, Е.В. Жолдыбаева

Подбор эффективного метода выделения ДНК для выявления сальмонелл методом полимеразной цепной реакции

При использовании молекулярно-биологических методов выделение ДНК является определяющим этапом, так как от качества и количества полученных препаратов ДНК во многом зависит результат ПЦР-анализа. В статье сравнили эффективность выделения ДНК бактерии рода *Salmonella* различными способами, а именно: триольный метод с использованием коммерческого реагента TRizol, СТАБ метод и сорбентный метод с использованием частиц кремнезема. Показано, что при выделении ДНК на частицах кремнезема обеспечивается высокая чувствительность выявления ДНК бактерии рода *Salmonella* по сравнению с другими методами. Амплификация экстрагированной ДНК на частицах кремнезема является более эффективной и позволяет получать воспроизводимые результаты высокой степени точности. В перспективе данный метод будет использован для комплектации тест-системы для диагностики сальмонелл. Внедрение методов экспресс-диагностики в практику чрезвычайно важно, так как быстрая и ранняя диагностика позволит своевременно противодействовать распространению инфекции, что, в свою очередь, повысит уровень эпидемиологической и эпизоотологической ситуации в стране.

Ключевые слова: бактерии *Salmonella*, метод, идентификация, выделение ДНК, полимеразная цепная реакция, амплификация, частицы кремнезема.

References

- 1 Tokarev, V.A., Gudkova, E.I., & Adarchenko, A.A. (1999). Variants monitorinha hospitalnykh shtammov VBI salmonelleznoi etiolohii [Variants of monitoring hospital strains of nosocomial infections, *Salmonella* etiology]. Proceedings of Nosocomial infections — problems of epidemiology, clinic, diagnosis, treatment, and prevention. Rossiiskaia nauchno-prakticheskaiia konferentsiia — Russian scientific-practical conference (pp. 236–237) [in Russian].
- 2 Lim, J.K., Gunther, N.W., Zhao, H., Johnson, D.E., Keay, S.K., & Mobley, H.L.T. (1998). In Vivo Phase Variation of *Escherichia coli* Type 1 Fimbrial Genes in Women with Urinary Tract Infection. *Infection and immunity*, 66, 7, 3303–3310.
- 3 Abzhaliyeva, A.B. (2017). Usovershenstvovanie metodov identifikatsii pishchevykh patogenov v produktakh zhivotnogo proiskhozhdeniya [Improving the identification of foodborne pathogens in animal products]. *Extended abstract of doctor's thesis*. Almaty [in Russian].
- 4 Nurgalieva, M.T., Zhanserkenova, O.O., Usenbekov, E.S., Kasymbekova, Sh.N., & Smagulov, A.K. (2016). Metody vydelenii nukleinovykh kislot dlja provedeniia polimeraznoi tseplnoi reaktsii v rezhime realnogo vremeni [Nucleic acid isolation methods for real-time polymerase chain reaction]. *Izdenister, natizheler — Research results*, 3, 71, 68–72 [in Russian].
- 5 Mamaev, M.A. (2000). Razrabotka uskorennyykh sposobov DNK-dianostiki dlja opredelenii bakterii v miase i miasoproduktakh [Development of accelerated methods of DNA diagnostics for the detection of bacteria in meat and meat products]. *Extended abstract of candidate's thesis*. Moscow [in Russian].

-
- 6 Herrera-Leo, S., McQuiston, J.R., & Usera, M.A. (2004). Multiplex PCR for Distinguishing the Most Common Phase-1 Flagellar Antigens of *Salmonella* spp. *J. Clin. Microbiol.*, 42, 2581–2586.
- 7 Scott O. Rogers, Arnold J. Bendich (1985). Extraction of DNA from milligram amounts of fresh, herbarium and mummified plant tissues. *Plant Molecular Biology*, 5, 69–76.
- 8 Kapustin, D.V., Prostiakova, A.I., & Alekseev, Ya.I. (2014). Vysokoeffektivnyi metod odnostadiinogo vydeleniya DNK dlja PTSR-diagnostiki Mycobacterium tuberculosis [Highly effective single-stage DNA extraction method for PCR diagnostics of Mycobacterium tuberculosis]. *Acta naturae*, 2, 21, 52–57 [in Russian].
- 9 Kirkimbaeva, Zh.S., Ermagambetova, S.E., & Biiashev, K.B. (2016). Sravnitelnoe izuchenie effektivnosti razlichnykh metodov vydeleniya DNK leptospir [A comparative study of the effectiveness of various methods of DNA isolation leptospiro]. *Izdenister, nauchnye — Research results*, 4, 72, 50–54 [in Russian].
- 10 Aukenov, N.E., Masabaeva, M.R., & Khasanova, U.U. (2014). Vydelenie i ochistka nukleinovykh kislot. Sostoianie problemy na sovremennom etape [Isolation and purification of nucleic acids. The state of the problem at the present stage]. *Nauka i zdravookhranenie — Science and Health*, 1, 51–53 [in Russian].
- 11 Kataloh saita «DNK-Tekhnologii» [Site Catalog «DNA-Technology»]. docplayer.ru. Retrieved from <https://docplayer.ru/45703435-I-ekspress-metod-vydeleniya-dnk-komplekt-reagentov-dlya-vydeleniya-dnk-proba-rapid-4-komplekt-reagentov-dlya-vydeleniya-dnk-proba-rapid-genetika-5.html> [in Russian].